

DOI: https://doi.org/10.70818/taj.v38i02.0363

Microbiological Profile and Antimicrobial Susceptibility Pattern on Blood Culture Isolates Enrolled in Microbiology Department at Rajshahi Medical College

Mizanur Rahman^{*1}, Kh Md Faisal Alam¹, Haimanti Shukla Das², Mahmuda Naznin¹, Quazi Tamanna Haque¹, Sahanaj Parvin¹

1 Department of Microbiology, Rajshahi Medical College, Rajshahi 2 Department of Virology, Rajshahi Medical College, Rajshahi

ABSTRACT: Background: Blood stream infections (BSI) are a significant global health

concern, contributing to both mortality and morbidity. These infections can range from

mild to life threatening, often requiring antimicrobial treatment. Due to increasing antimicrobial resistance and changing patterns of antibiotic use, the epidemiology and

outcomes of BSI are constantly evolving. Therefore, continuous monitoring of the

bacterial causes of BSI and their antibiotic resistance pattern is necessary. Objective: To

find out the bacteriological profile and their antibiotic sensitivity patterns among

suspected BSI patients. Materials and Methods: A cross-sectional study was carried out

in the Department of Microbiology, Rajshahi medical college from January 2024 to

December 2024. A total of 1228 suspected BSI patients were included in the study. The

BacT/Alert automated blood culture method was used to isolate bacterial pathogens and antimicrobial susceptibility test was performed by modified Kirby-Bauer disc diffusion

method following CLSI 2024 guidelines. Results: Out of 1228 cases, 125 (10.17%) were

culture positives, where male were 72(57.6%) and female were 53(42.4%). Among 125

culture positive cases, 86 (68.8%) were Gram negative bacteria and 39 (31.2%) were Gram positive bacteria. The most prevalent pathogens were *Coagulase negative staphylococcus*

29(23.2%) and E. coli 27(21.6%). Gram negative bacteria are resistant to Amoxyclav, 2nd

and 3rd generation Cephalosporin and lower resistance was shown to Meropenem,

Piperacilin/tazobactam and Amikacin. Conclusion: Regular monitoring of sensitivity

patterns, creating hospital antibiotic policies based on current data and following

treatment guidelines can encourage appropriate antibiotic use and reduce bacterial

Keywords: Antibiotic-Resistance, Antibiotic Sensitivity, Bacterial Isolates, Blood Culture,



Citation:

Rahman M, Alam KMF, Das HS, Naznin M, Haque QT, Parvin S; Microbiological Profile and Antimicrobial Susceptibility Pattern on Blood Culture Isolates Enrolled in Microbiology Department at Rajshahi Medical College. Journal of Teachers Association. 2025;38(2): 273-279

Article History:

Received: 03.02.2025 Accepted: 18.04.2025 Published: 01.06.2025



Copyright © 2025 The Author(s): This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC BY-NC 4.0) which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.

Article at a glance:

Study Purpose: To contribute to existing knowledge or propose new ideas.

resistance.

Bloodstream Infections.

Key findings: Among Gram negative bacteria, E. coli 27 (21.6%) and among Gram positive bacteria CoNS 29(23.2%) were identified as most common isolates.

Newer findings: In this study, Gram negative bacteria were showed higher resistance to commonly used antibiotics from BSIs.

INRODUCTION

Blood stream infections (BSI) are a significant cause of mortality and morbidity globally, ranging from self-limiting to life threatening sepsis requiring antimicrobial treatment.¹Bacteremia is the presence of viable bacteria in the blood while septicemia indicates systemic manifestations caused by bacteria or their toxins in blood. Hospital-acquired BSIs range from 9% to 11% in developed nations and up to 19% in low and middle countries.^{2, 3} Factors contributing to BSIs include medical devices, patient age and pre-existing medical condition such as diabetes mellitus, cancer, renal failure, burns and previous hospitalization.⁴ Other risk variables such as the severity, age, sex all affects the death rate from bloodstream infections.⁵ Blood culture remains gold standard for diagnosing blood stream infections, despite bacteria are not always being identified.⁶ Multidrug resistance especially in Gram negative bacteria causing BSIs, poses a significant therapeutic challenge, leading to fewer treatment options, higher cost, longer hospital stay and increased morbidity and mortality.⁷ Antibiotic-resistance varies geographically, necessitating surveillance and documentation of blood culture isolates to guide the best empirical antibiotic use and reduce resistance.⁸ With this above view, to isolate and identify different bacterial causes of BSIs, determining the antibiotic susceptibility patterns of isolated bacteria and suggesting empirical treatment of BSIs.

MATERIALS AND METHODS

This cross-sectional descriptive study was conducted in the Rajshahi Medical College's Microbiology department over a period from January to December 2024. The study involved collecting two venous blood samples were taken from two separate locations, 30 minutes apart, from each participant who had a suspected BSIs following strict aseptic procedures. The sample sets consisted of 8-10 ml of adult venous blood, 0.5- 2 ml of neonatal venous

RESULTS

A total of 1228 blood culture samples were collected from clinically suspected patients with BSI during the study period. Only 125 (10.17%) of the 1228 samples tested positive for culture, whereas 1103 (89.83%) tested negative (Figure 1). Among the culture 125 positive cases, higher in males (72; 57.6%) as compared to females (53; 42.4%) (Figure 2). Of the 125 bacterial isolates, 86 (68.8%) were Gram-negative bacteria, while the remaining 39(31.2%) were Grampositive bacteria (Table 1). Amongst the Gramnegative bacteria, most isolated organism was E. coli (21.6%) followed by Klebsiella spp. (19.2%), blood or 2-5 ml of blood from pediatric patients. These sample were immediately inoculated into aerobic blood culture bottles (adult or pediatric bottles, depending on the situation) and incubated using BacT/ALERT® 3D automated blood culture analyzer. All broths that tested positive for BacT/ALERT were Salmonella subcultured onto Shigella agar, MacConkey's agar and blood agar. Subculturing onto blood agar and MacConkey's agar was carried out on days 2, 5, and 7 days of incubation for those bottles without positive signs. For 18 to 24 hours, the inoculated blood agar, MacConkey's agar, and Salmonella Shigella agar plates were incubated at 37 °C. Using standard laboratory procedures, colony morphology, gram staining, and traditional biochemical testing were used to identify bacterial growth. 9 The modified Kirby-Bauer disc diffusion method assessed the antibiotic susceptibility of isolated bacteria on Mueller-Hinton agar with CLSI guideline, determining if the results were sensitive or resistant. 10

Pseudomonas spp. (14.4%), *Salmonella spp* (12%) and *Acinetobacter spp.* (1.2%) respectively. Antimicrobial susceptibility pattern of Gram-negative bacteria showed highest susceptibility to Cefepime, Meropenem, Piperacillin/tazobactam and lowest susceptibility to Amoxyclav, Cefuroxime, Cefixime and Ceftriaxone (Table 2). The susceptibility pattern of Gram-positive isolates showed highest susceptibility to Vancomycin, Linezolid, Meropenem and lowest susceptibility to Azithromycin, Ciprofloxacin (Table 3).

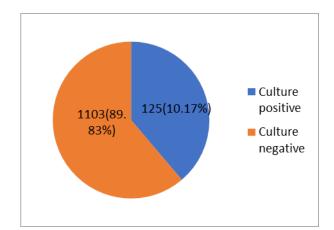


Figure 1: Frequency of culture positive and culture negative cases (N=1228)

Mizanur Rahman et al.; Journal of Teachers Association, Apr-Jun, 2025; 38(2): 273-279

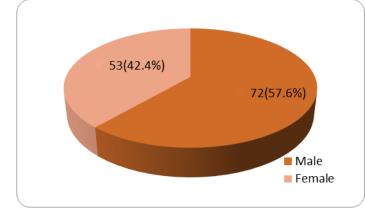


Figure 2: Gender variation regarding growth positive cases: (n=125)

Table 1: Gram positive and Gram-negative bacteria among bacterial growth (n=125)Isolated Organism Number (%)

Gram negative	86(68.80%)
Gram positive	39(31.20%)

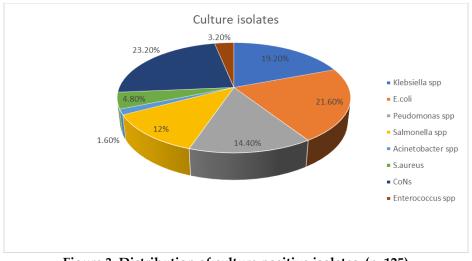


Figure 3: Distribution of culture positive isolates. (n=125)

Table 2: Antimicrobial	susceptibility pattern	of Gram-negative of	organism. (n=86)

Antimicrobial		E.coli	Klebsiella spp.	Pseudomonas spp.	Salmonella spp.	Acinetobacter spp.
Agent		(27)	(24)	(18)	(15)	(02)
Amikacin	S	70.83%	81.49%	66.67%	66.67%	100%
	R	29.17%	18.51%	33.33%	33.33%	0%
Amoxyclav	S	62.50%	88.89%	61.11%	80%	-
	R	37.5%	11.11%	38.89%	20%	-
Aztreonam	S	70.83%	48.14%	72.22%	-	-
	R	29.17%	51.86%	27.78%	-	-
Azithromycin	S	-	-	61.11%	26.67%	-
	R	-	-	38.89%	73.33%	-
Ampicillin	S	-	-	-	86.67%	-
	R	-	-	-	13.33%	-
	S	37.5%	74.08%	-	66.67%	-

© 2025 TAJ | Published by: Teachers Association of Rajshahi Medical College

				Mizanur Rahman et al.; J	ournal of Teachers Associat	tion, Apr-Jun, 2025; 38(2): 273-279
Cefixime	R	62.5%	25.92%	-	33.33%	-
Cefepime	S	70.83%	70.38%	-	60%	50%
	R	29.17%	29.62%	-	40%	50%
	S	33.33%	81.49%	-	80%	0%
Ceftriaxone	R	66.67%	18.51%	-	20%	100%
	S	37.5%	25.92%	-	-	-
Cefuroxime	R	62.5%	74.08%	-	-	-
Cotrimoxazole	S	-	-	-	80%	0%
	R	-	-	-	20%	100%
Ciprofloxacin	S	66.67%	29.62%	66.67%	73.33%	100%
	R	33.33%	70.38%	33.33%	26.67%	0%
Ceftazidime	S	66.67%	44.44%	72.22%	-	50%
	R	33.33%	55.56%	27.78%	-	50%
Colistin	S	-	-	-	-	100%
	R	-	-	-	-	0%
Meropenem	S	75%	66.67%	83.33%	100%	100%
-	R	25%	33.33%	16.67%	0%	0%
Netilmicin	S	-	-	66.67%	-	-
	R	-	-	33.33%	-	-
Piperacillin/	S	83.33%	48.14%	83.33%	-	100%
tazobactam		16.67%	51.86%	16.67%	-	0%
	R					
Levofloxacin	S	-	-	-	100%	-
	R	-	-	-	0%	-

Table 3: Antimicrobial susceptibility pattern of Gram-positive bacteria (n=39)

Antimicrobial agent		Enterococcus spp. (04)	CoNS (29)	Staphylococcus aureus (06)
Amikacin	S	75%	89.65%	66.67%
	R	25%	10.34%	33.33%
Amoxyclav	S	50%	62.06%	33.33%
	R	50%	37.94%	66.67%
Azithromycin	S	50%	27.58%	33.33%
	R	50%	72.41%	66.67%
Cotrimoxazole	S	50%	55.17%	50%
	R	50%	44.82%	50%
	S	75%	37.93%	66.67%
Ciprofloxacin	R	25%	62.07%	33.33%
Linezolid	S	100%	100%	83.33%
	R	0%	0%	16.67%
Meropenem	S	75%	81.10%	66.67%
	R	25%	18.90%	33.33%
Vancomycin	S	100%	100%	100%
	R	0%	0%	0%

DISCUSSION

Blood culture is a well-established procedure of the standard diagnostic workup for many infectious diseases. In Bangladesh, all kinds of drugs including the antibiotics are sold over the counter, misuse of antibiotics has been found to be responsible for developing pool of resistant bacteria as well as negative results of blood culture.¹¹ A total of 1228 blood culture samples were collected from clinically suspected patients with BSI during the study period. Only 125 (10.17%) of the 1228 samples tested for culture positive, whereas 1103 (89.83%) tested negative (Figure 1). This study were nearly similar with Laxmi Kant Khanal et al., in Nepal; Nasrin et al., in Bangladesh with 10.3%, 13% were cultures positive and 89%, 87% were found culture negative respectively.^{12,13} The current findings were dissimilar with Shewta et al., in India; Daniel Ningthoujam et al., in India with culture positive 25.8%, 31.2% and 74.2%, 68.8% were culture negative.14,15 The variation in positivity rates among different studies may be attributed to differences in the methodology used for blood culture, the volume or number of blood culture samples taken, study design, geographical differences, nature of the patient population, differences in the epidemiological agents and variations in infection control policies.¹⁵ Among the 125 culture positive cases, higher in males (72; 57.6%) as compared to females (53; 42.4%) (Figure 2). The finding corresponded with the study of Belew et al., in Ethiopia (50.9% male and 49.1% female) and Shewta et al., in India (male 59% and female 41%).14,20 Dissimilar with R.S. Parihar et al., in India (male 68.75% and female 31.25%).¹ This may be explained as men are involved in more physical activities for livelihood and less frequent hand hygiene practice which could potentially provide environment for large reservoirs of common pathogens responsible for causing blood stream infections.¹ Of the 125 bacterial isolates, 86 (68.8%) were Gram-negative bacteria, while the remaining isolates 39(31.2%) were Grampositive bacteria (Table 1). In contrast to this study, similar findings were found with 68% Gram negative and 32% Gram positive bacteria of Habyarimana et al., in Africa and 60% Gram negative and 40% Gram positive were found of Laxmi Kant Khanal et al., in Nepal.^{12, 18} Dissimilarity was found with Daniel Ningthoujam et al., in India and Cheema K H et al., in Pakistan which were found 24.7% and 96.9% were Gram negative bacteria and 75.3% and 3.1% as Gram positive bacteria respectively.15,19

Among the Gram-negative bacteria (Figure 3), most isolated organism was *E.coli* (21.6%) followed by *Klebsiella spp* (19.2%), *Pseudomonas spp* (14.4%), *Salmonella spp* (12%) and *Acinetobacter spp* (1.6%) respectively. Among Gram positive 39 cases, highest organism *CoNS* (23.2%) followed *S. aureus* (4.8%) and *Enterococcus spp* (3.2%) (Figure 3). Among the Gramnegative bacteria, the similar study were found *E. coli* (29.4%), *Klebsiella spp* (20.6%), *Pseudomonas spp* (8.8%) with Oluwalana T *et al.*, in Nigeria and *E.coli* (14%), *Klebsiella spp*. (13%), *Acinetobacter spp* (7%), *Pseudomonas spp*. (7%) with Kaur C and Sharma

S, in India.¹⁶,²⁰ Dissimilar were found E.coli (5.7%), Klebsiella spp. (5.7%), Acinetobacter spp. (7%), Pseudomonas spp. (1.4%) Salmonella spp. (59.5%) with Cheema K H et al., in Pakistan and Pseudomonas spp (11.3%), Acinetobacter spp. (6%), Klebsiella spp (6%), E. coli (0.7%), Salmonella spp 1 (0.7%) with Daniel Ningthoujam et al., in India.^{19,15} Figure 3 showed that among 39 Gram positive cases the highest organism was CoNS (23.2%) followed S. aureus (4.8%) and Enterococcus spp. (3.2%). Similar with Kaur C and Sharma S., in India were found CoNS (31.2%) S. aureus (7%) followed by Enterococcus spp (6%) and Laxmi Kant Khanal, in Nepal were found CoNS (26.8%) and Enterococcus spp (6.52%).^{20,12} Different dissimilar were found with R.S. Parihar et al., in India, Oluwalana T et al., in Nigeria, where CoNS were (41.3%, 8.8%) and S. aureus was (23.8%) respectively.^{16, 17}

Antimicrobial susceptibility pattern of Gramnegative bacteria showed highest susceptibility to Cefepime, Meropenem, Piperacillin/tazobactam and lowest susceptibility to Amoxyclav, Cefuroxime, Cefixime and Ceftriaxone (Table 2). E. coli was the most susceptible to Meropenem (75%), Amikacin Piperacillin-tazobactam (70.83%) and (83.33%)whereas Cefixime (62.5%) and Ceftriaxone (66.66%) were the least susceptible drug. This study similar to Shewta et al., in India for Piperacillin-tazobactam (81.81%), Meropenem (81.81%) and Amikacin (72.72%) & Zerin T et al., in Bangladesh Meropenem (84.57%), Amikacin (60.64%) and Cefixime (60%), Ceftriaxone (44.68%), respectively had the lowest susceptibilities.14, 21

Among the Gram-negative isolates, Klesiella showed highest susceptibility spp towards Meropenem (66.67%), Amikacin (81.49%) and Cefepime (70.38%) and lowest susceptibility to Amoxyclav (11.11%), Ceftriaxone (18.51%), Cefixime (25.92%) respectively. Similar study was found Belew et al., in Ethiopia in highest susceptibility to Meropenem (62.5%), Amikacin (78.5%) and lowest susceptibility Amoxyclav (0%) and Ceftriaxone (18.51%) respectively.²¹ Dissimilar with Cheema K H et al., in Pakistan was found to Meropenem (40%), Amikacin (40%) and lowest susceptibility Amoxyclav (0%) and Ceftriaxone (30%).¹⁹ and R.S. Parihar et al., in India was found Meropenem (100%), Amikacin (57.1%) respectively.¹

Table 3 *CoNS* had the lowest susceptibility to Azithromycin (72.41%) and Ciprofloxacin (62.06%) among Gram-positive isolates, whereas the highest susceptibility to Vancomycin (100%), Linezolid (100%) and Meropenem (81.1%) was observed. Similar studies were conducted in India by R.S. Parihar *et al.*, in with 100% susceptible to Vancomycin and Linezolid and Shewta *et al.*, in with 100% Vancomycin and 92.85% Linezolid.^{1, 14} The three antibiotics that *S aureus* was most susceptible to were Vancomycin (100%), Linezolid (83.33%), and Meropenem (66.67%). Similar studies were conducted in India by Shewta *et al.*, in with Vancomycin (100%) and Linezolid (80%) and R.S. Parihar *et al.*, in with Vancomycin (100%) and Linezolid (94.73%).^{14, 1}

CONCLUSION

The resistance pattern of Gram-negative bacilli to commonly used drugs has raised concerns among clinicians and hospital groups to treat blood stream infections. So, detection of bloodstream infection-causing pathogens and their antibiogram which can help choose the best empirical treatment.

Ethical Approval

Ethical clearance for the study was taken from the Instutional Review Board and concerned authority, Rajshahi Medical College & Hospital.

Conflict of Interest: None declared.

Funding: No funding sources.

Consent: Informed written consent was taken from each patient's attendant.

REFERENCES

- 1. R.S. Parihar et al. Bacteriological Profile and Antimicrobial Susceptibility Patterns of Blood Borne Pathogens in a Tertiary Care Center, Jodhpur (Rajasthan), India. Int. J. Curr. Microbial. App. Sci. 2018; 7(10): 1785-1792.
- 2. Report on the burden of endemic health careassociated infection worldwide. Geneva: World Health Organization; 2011.
- Suetens C, Latour K, Karki T, et al. Prevalence of healthcare-associated infections, estimated incidence and composite antimicrobial resistance index in acute care hospitals and long-term care facilities: Results from two European point prevalence surveys, 2016 to 2017. Euro Surveill. 2018; 23(46):1800516.

- 4. Sainfer A, Bevin C, Jianfang L, et al. Prevalence and risk factors for bloodstream infections present on hospital admission. J Infect Prev. 2018;19(1):37–42.
- 5. Christaki E, Giamarellos-Bourboulis EJ. The complex pathogenesis of bacteremia: From antimicrobial clearance mechanisms to the genetic background of the host. Virulence. 2014;5(1):57–65.
- 6. Paolucci M, Landini MP, Sambri V. How can the microbiologist help in diagnosing neonatal sepsis? Int J Pediatr. 2012; 2012:120139.
- 7. Howard DH. The global impact of drug resistance. Clin Infect Dis. 2003;36: S4–S10.
- Jadhav S, Gandham N, Paul R, et al. Bacteriological profile of septicaemia and antimicrobial susceptibility of isolates from tertiary care hospital in India. Res J Pharm Biol Chem Sci. 2012;3(4):1100–1108.
- Collee, J.G., Miles, R.S. and Watt, B. Tests for the Identification of Bacteria. In: Collee, J.G., Marmion, B.P., Fraser, A.G. and Simmons, A., Eds., Mackie & McCartney Practical Medical Microbiology, 14th Edition, Churchill Livingstone, New York, 1996; p. 131–151.
- CLSI (2024). Performance Standards for Antimicrobial Susceptibility Testing. 34th ed. CLSI supplement M100.Wayne, P.A.: Clinical and Laboratory Standards Institute.
- 11. 11.Wadud A, Khali MI, Shamsuzzaman A, et al. Bacteriological profiles of Blood culture isolates by BacT/ALERT 3D automated system. Journal of Shaheed Suhrawardy Medical College. 2009;1(2).
- 12. Khanal LK. Bacteriological Profile of Blood Culture and Antibiogram of the Bacterial Isolates in a Tertiary Care Hospital. International Journal of Health Sciences and Research.2020;10(8).
- 13. Nasrin M, Begum FM, Karim R, et al. Bacteriological profile and antimicrobial susceptibility patterns of blood culture isolates among bloodstream infection suspected patients attending in a referral hospital.2021; 15 (2): 5-11.
- 14. Shweta et al. A study on microbiological profile and antimicrobial resistant pattern of blood culture isolates in critical care units in a tertiary care hospital in North India. 2024; 6 (5); 244-248.
- 15. Daniel Ningthoujam et al. Bacteriological Profile and Antibiogram of Aerobic Blood Culture Isolates among Suspected Blood Stream Infections: A Cross-sectional Study. National Journal of Laboratory Medicine. 2024;13(1).

© 2025 TAJ | Published by: Teachers Association of Rajshahi Medical College

- 16. Oyekale OT, Ojo BO, Olajide AT, et al. Bacteriological profile and antibiogram of blood culture isolates from bloodstream infections in a rural tertiary hospital in Nigeria. 2022; 11(1), 1807.
- 17. Habyarimana et al. Bacteriological Profile and Antimicrobial Susceptibility Patterns of Bloodstream Infection at Kigali University Teaching Hospital. Infection and Drug Resistance 2021:14 699–707.
- 18. Cheema KH, Anwar MS, Hameed F, et al. Assessment of Bacterial Profile and Antimicrobial Susceptibility Pattern of Blood Culture Isolates. JIIMC 2022; 17(1), 9-11.
- 19. Kaur C, Sharma S. Bacteriological Profile and their Antibiotic Susceptibility Pattern in

Bloodstream Infections in a Tertiary Care Hospital in North India. 2022;16(4):2756-2763.

- 20. Belew et al. Phenotypic Bacterial Isolates, Antimicrobial Susceptibility pattern and Associated factors among Septicemia Suspected Patients at a hospital, in Northwest Ethiopia: Prospective cross-sectional study. Annals of Clinical Microbiology and Antimicrobials. RESEARCH (2023) 22:47.
- 21. Zerin T *et al.*, Identification and Antibiotic Susceptibility of Blood Culture Isolates from Rajshahi, Bangladesh Journal of Scientific Research in Medical and Biological Sciences. ISSN 2709-0159 and 2709-1511:2021;2(2).

*Correspondence: Dr. Mizanur Rahman, Email: zarif345@yahoo.com

Journal of Teachers Association Official Journal of Teachers Association Rajshahi Medical College



Publish your next article in TAJ For submission scan the QR code E-mail submission to: tajrmc8555@gmail.com